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W-8000 München 86(DE)**(54) **DNA coding for human cell surface antigen.**

(57) DNAs coding for human cell surface antigen (Fas or Fas antigen), vectors for expressing for said DNAs and transformants transfected with said vector are provided. Fas is a polypeptide that exists in the surfaces of a variety of cells and is considered to be deeply concerned with the apoptosis of cells. The isolated Fas cDNA has an open reading frame that is capable of encoding a protein consisting of 335 amino acids. The mature Fas antigen is a protein consisting of 319 amino acids having a calculated molecular weight of about 36,000 and is constituted by an extracellular domain of 157 amino acids, a membrane-spanning domain of 17 amino acids, and a cytoplasmic domain of 145 amino acids.

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The present invention relates to DNAs coding for human cell surface antigen (hereinafter referred to as Fas or Fas antigen) and to vectors for expressing for said DNAs.

Fas is a polypeptide that exists in the surfaces of a variety of cells and is considered to be deeply concerned with the apoptosis of cells. The apoptosis is a form of death of cells that is distinguished from the so-called necrosis of cells, and is observed at the time of death of various cells such as of embryogenesis, metamorphosis, endocrine-dependent tissue atrophy and turnover of normal tissues [Wyllie et al. *Int. Rev. Cytol.* 68, 251-306, 1980; Walker et al. *Meth. Achiev. Exp. Pathol.* 13, 18-54, 1988; Schmidt et al. *Proc. Natl. Acad. Sci. USA* 83, 1881-1885, 1986; Ucker et al. *Nature* 327, 62-64, 1987; Smith et al. *Nature* 337, 181-184, 1989, Williams et al. *Nature* 343, 76-79, 1990]. The following features have been pointed out as a result of the morphological and biochemical analyses of cells at the apoptosis:

The apoptosis is accompanied by condensation of cytoplasm, loss of plasma membrane microvilli, segmentation of the nucleus, extensive degradation of chromosomal DNA (into oligomers of about 180 base pair units), and formation of apoptotic bleb [Wyllie et al. 1980 (mentioned above)]. The apoptosis is a physiologically and medically interesting phenomenon because it is a form associated with the death of immunocytes such as thymocytes and the extinction of the tumor cells.

In regression of tumor (alleviation of tumor), in general, the apoptosis mediates the death of target cells by interaction with natural killer cells or cytotoxic T lymphocytes [Duke et al. *Proc. Natl. Acad. Sci. USA* 80, 6361-6365, 1983; Schmidt et al, 1986 *ibid.*; Ucker, 1987 (mentioned above)], or by tumor necrosis factor- α (TNF- α) or its related cytokine lymphotoxin (TNF- β) against the target cells [Schmidt et al, 1986 (mentioned above); Dealtry et al. *Eur. J. Immunol.* 17, 689-693, 1987; Larrick and Wright, *FASEB J.* 4, 3215-3223, 1990].

With regard to the relationship between the Fas antigen and the apoptosis, the present inventors have previously disclosed that the mouse monoclonal antibody against the human Fas antigen has a cytolytic activity on human cells expressing the Fas antigen while it does not act upon mouse cells [Yonehara et al. *J. Exp. Med.* 169, 1747-1756, 1989]. It has also been disclosed by Trauth et al. that the anti-Apo-1 antibody has effects analogous to those of the anti-Fas antibody called Anti-Apo-1 [Science 245, 301-305, 1989].

In a recent study by the present inventors, furthermore, it has been found that cells infected with human immunodeficiency virus (HIV) are more sensitive to the cytotoxic activity of the anti-Fas monoclonal antibody than uninfected cells [Kobayashi et al. *Proc. Natl. Acad. Sci. USA* 87, 9620-9624, 1990]. However, it is still not clear whether the expression of the Fas antigen that is predominant in the infected cells is actually induced by infection with HIV or by a general transformation. It is also considered potential to specifically drive the HIV-infected cells into apoptosis by using a monoclonal antibody specific to Fas.

The present inventors have further discovered that the treatment of human colon carcinoma HT-29 cells with interferon- γ (INF- γ) induces the Fas antigen on the cell surface, and renders the tumor cells more susceptible to the cytotoxic activity of the anti-Fas antibody [Yonehara et al, 1989 (mentioned above)].

As described above, it has been pointed out that the Fas antigen is closely related to the apoptosis but numerous points remain not clarified. Therefore, it is physiologically and pathologically meaningful to disclose the entire structure of the Fas antigen and to clarify its function. It is further considered that various monoclonal antibodies that specifically reacts with Fas may be easily obtained if the structure of the Fas antigen is disclosed, and used in treating diseases associated with HIV infection and malignant tumors to be cured.

In order to accomplish the above object, it is essential to establish means capable of supplying Fas polypeptides in sufficient amounts. In recent years, a recombinant DNA technology has been utilized as a method for preparing physiologically active substance. In order to prepare the Fas antigen by utilizing the above technology, however, it is necessary to isolate DNA that encodes Fas proteins followed by cloning.

The present inventors have succeeded in the development of means capable of producing in large amounts the human Fas antigen in pure form. The present inventors have clarified the genes of the human Fas antigen and have disclosed, for the first time, how to genetically manipulate the Fas antigen genes. The present invention provides DNA coding for human Fas antigens, DNA derived therefrom, and DNA fragments thereof. They may include those having an anti-sense sequence thereof. The present invention further provides products such as proteins and peptides produced by using the DNA that encodes the Fas antigen or by using derivatives thereof.

The invention also provides plasmids or vectors that carry DNA coding for the Fas antigen or DNA derived therefrom or fragments thereof. Moreover, the invention provides a variety of transformants that hold replicably or expressibly the plasmid or the vector therein. The present invention encompasses a variety of products produced by utilizing base sequence information of DNA encoding the Fas antigen.

Fig. 1 shows the nucleotide sequence and predicted amino acid sequence of the cDNA coding human Fas protein (up to 284th amino acid).

Fig. 2 shows the nucleotide sequence and predicted amino acid sequence of the cDNA coding human Fas protein (after 284th amino acid).

Fig. 3A shows the schematic representation and restriction map of the human Fas cDNA (pF58).

Fig. 3B shows the hydropathy plot of amino acid sequence of human Fas antigen.

5 Fig. 4 shows the graph representing the results examined by a flow fluorometry for the expression of the human Fas antigen in mouse cells transformed with the human Fas expression vector.

Fig. 5 shows the graph representing cytolytic effect of the anti-Fas antibody on the WR19L transformant clones.

10 Fig. 6 shows the graph representing cytolytic effect of the anti-Fas antibody on the L929 transformant clones.

Fig. 7 shows the schematic representation of comparison in amino acid sequence of Fas antigen with other members of the NGFR/TNFR family.

Fig. 8 shows the schematic representation of comparison in amino acid sequence of extracellular domain of the human Fas with other members of the NGFR/TNFR family.

15 Fig. 9 shows the comparative representation of the amino acid sequences of the cytoplasmic domains of the Fas, TNF receptor type I and CD40.

The invention relates to DNA coding for human cell surface antigen or those having substantially the same functions as said human cell surface antigen, DNA derived therefrom or DNA fragmented therefrom. Particularly, the invention relates to DNA coding for Fas antigens, preferably peptides having at least a part of the amino acid sequences, and more preferably the amino acid sequences described in Figs. 1 and 2.

20 Furthermore, the invention relates to DNA comprising at least a part of the base sequences described in Figs. 1 and 2, preferably DNA having the base numbers 215 to 1199, 243 to 1199, 215 to 713 or 243 to 713 of Figs. 1 and 2, or a portion thereof.

The invention still relates to proteins or peptides comprising at least a part of the amino acid sequences having a substantially human cell surface antigen activity, particularly a Fas antigen activity, preferably at least a part of the amino acid sequences described in Figs. 1 and 2, and more preferably the amino acid numbers -16 to 319, 1 to 319, -16 to 157, or 1 to 157 described in Figs. 1 and 2.

25 The invention also relates to expression vectors comprising the above DNA, transformants transformed by said expression vector and methods for producing said protein or peptide which comprises cultivating said transformant under a suitable condition in a suitable medium and collecting the produced protein or peptide from the cultured medium.

30 The present invention is also concerned with various reagents for analysis or medical drugs comprising an effective amount of the product such as proteins obtained as described above as well as antigens obtained as described above.

35 According to the present invention, it would become possible to develop Fas genes or Fas gene analogs in various cells inclusive of human cells by utilizing information related to base sequences of the cDNA clone (for example, pF85) or fragments derived therefrom or base sequences thereof.

It should be comprehended that the present invention is concerned with those that are thus finally obtained. The development can be effected according to methods described in this specification or according to suitably modified methods.

40 The present inventors have screened a variety of human cell lines in connection with the expression of the as antigen and have discovered that human T cell lymphoma KT-3 expresses the Fas antigen about 20 times as much as other cell strains. The inventors have succeeded in isolating and cloning cDNAs encoding human Fas antigen determinant from human T cell lymphoma KT-3 cells.

45 FIGs. 1 and 2 show a cDNA nucleotide sequence and predicted amino acid sequence from a human Fas antigen cDNA clone (pF 58) that is obtained herein.

Fig. 3 shows a restriction map of cDNA (pF 58) for human Fas antigen.

50 The transformant (*Escherichia coli*, pF 58) carrying the plasmid pF 58 was originally deposited as a domestic microorganism deposit (FERM P-12192) at the Fermentation Research Institute, Agency of Industrial Science and Technology, Ministry of International Trade and Industry, Japan (FRI) on April 12, 1991 and converted into an international one (FERM BP-3826) under the Budapest Treaty.

The above pF58 cDNA has an open reading frame that is capable of encoding a protein consisting of 335 amino acids. From the predicted amino acid sequence, it is estimated that the mature Fas antigen is a protein consisting of 319 amino acids and is constituted by an extracellular domain, a transmembrane domain and a cytoplasmic domain. Such a constitution is common to many cell surface receptors. As will be described later, it was confirmed through the comparison of the amino acid sequence of the Fas protein with amino acid sequences of other cell surface proteins that the above Fas protein pertains to an NGFR/TNFR family in the group of cell surface membrane proteins.

The cDNA (e.g. pF58) encoding the Fas antigen of the present invention is inserted into a plasmid for expression under the regulation of a human peptide chain elongation factor 1 α gene promoter to construct an expression plasmid (e.g. pEFF-58). According to the present invention, mouse T cell lymphoma WR19L and mouse fibroblastoma L929 cells are transformed with the above expression plasmid. The flow cytometry analysis of the transformants revealed that the Fas antigen is expressed in very large amounts on their surfaces. It has been further confirmed that the transformed cell lines exhibit a dose-dependent response to the anti-Fas antibody and die. Through the observation of morphological changes, fragmentation of chromosomes and the like, it has been made clear that these cells die due to apoptosis.

The present invention provides DNAs coding for human cell surface antigen Fas and expression vectors for carrying the DNA.

The cDNA (e.g. pF58) encoding the Fas antigen of the present invention can be isolated by ordinary methods from the transformant (e.g. *Escherichia coli*, pF 58 which carries the plasmid pF 58 was habored was originally deposited as a domestic microorganism deposit (FERM P-12192) at the Fermentation Research Institute, Agency of Industrial Science and Technology, Ministry of International Trade and Industry, Japan (FRI) on April 12, 1991 and converted into an international one (FERM BP-3826) under the Budapest Treaty.

The cloning of cDNA coding for the human Fas antigen according to the present invention can be carried out according to conventional methods in the art. The total RNAs are prepared from cells expressing human Fas antigen (e.g. KT-3 cell) and poly(A)RNAs are isolated. Then, a double stranded cDNA is synthesized by using reverse transcriptase or the like enzyme and is introduced into a mammal expression vector (e.g. pCEV4, Ito et al., 1990) to prepare cDNA libraries. The cDNA libraries (e.g. cDNA libraries of about 8×10^5 independent clones) are transfected into mammal cells (e.g. COS-7 cell) by the spheroplast fusion method or the like. After the transfection (e.g. at 72 hr posttransfection), the transfected mammal cells (e.g. the transfected COS-7 cells) are incubated with anti-Fas antibody (e.g. mouse anti-Fas antibody (IgM)), and the mammal cells expressing the Fas antigen (e.g. the COS cell expressing the Fas antigen) are recovered by the panning procedure [Seed and Aruffo, Proc. Natl. Acad. Sci. USA 84, 3365-3369, 1987] using goat anti-mouse IgM or the like.

The extrachromosomal DNA is prepared from the adherent mammal cells (e.g. the adherent COS cells) according to the method of Hirt [J. Biol. Chem. 264,14929-14934, 1967] or the like, and introduced into *Escherichia coli* or the like. The resultant colonies are pooled, used for spheroplast fusion, etc. with mammal cells (e.g. COS cell), and the panning is performed as described above. This procedure is repeated (e.g. three times) to obtain individual clones (e.g. 14 individual clones (pF1 to pF14)). Then, mammal cells (e.g. COS cells) are transfected with selected clones (e.g. pF1 having 3.0 kb insert and pF3 having 1.5 kb insert) among the individual clones. The resulting cells are analyzed by the flow cytometry using an anti-Fas antibody and the like. In a preferred embodiment of the present invention, it has been found that two cDNAs code for proteins that have the Fas antigen determinant. The pF1 and pF3 have been subjected to the restriction enzyme mapping and the DNA sequencing analysis. As a result, it has been found that the pF1 and pF3 share identical sequences at the 5' end including about 500 bases. However, their sequences at the 3' end diverge completely (see Fig. 3A).

Next, the original cDNA libraries of cells expressing human Fas antigen are screened by the colony hybridization using an isolated DNA fragment derived from cDNA coding for proteins related to the human Fas antigen (e.g. XhoI-BamHI DNA fragment at the 5' end of the pF3). As a result, clones which have full-length DNA encoding Fas antigen are obtained. In a preferred embodiment of the present invention, it has been found that ten clones are isolated and subjected to restriction enzyme mapping. These cDNAs contained inserts of 1.8 to 2.6 kb, showed identical restriction maps and overlapped each other. The longest cDNA clone (pF58) was selected from the resulting clones. Fig. 3 shows the restriction map of the longest cDNA clone (pF58), and Fig. 1 and 2 show the nucleotide sequence and the predicted amino acid sequence.

The pF58 cDNA has a long open reading frame of 1008 nucleotides capable of coding for a protein consisting of 335 amino acids. The hydropathy analysis of the predicted amino acid sequence indicates the presence of a signal sequence at the N-terminal end (Fig. 3B). Comparison of the N-terminal sequence with typical signal peptide cleavage sites suggests that the mature Fas antigen would lack the signal peptide portion and be a protein consisting of 319 amino acids having a calculated molecular weight of about 36,000. This Fas antigen protein consists of an extracellular domain of 157 amino acids, a membrane-spanning domain of 17 amino acids, and a cytoplasmic domain of 145 amino acids.

Western blotting analysis of the membrane fractions from KT-3 cells and the WR19L transformant clone, F58-12A, expressing the Fas antigen using the anti-Fas antibody, shows a specific band with an apparent molecular weight of about 43,000. This value is in good agreement with the above calculated value

(about 36,000) from the standpoint in which sugar moieties are bonded to two potential N-glycosylation sites found in the extracellular domain of the Fas antigen (see Fig. 1).

Moreover, the KT-3 cells are subjected to the northern hybridization using the Fas antigen cDNA or its fragment as a probe to detect two bands at 2.7 and 1.9 kb. By taking the presence of the poly(A) tail into consideration, it is considered that the larger mRNA is almost identical to the size of the above pF58 cDNA. It is therefore considered that pF58 is a full-length cDNA for the larger mRNA. If human colon carcinoma HT-29 cells are treated with 300 units/ml human INF- γ for 7 hours prior to harvest, both large and smaller mRNAs for the Fas antigen are expressed distinctly.

Forty percent of the cDNA clones isolated from the KT-3 cDNA libraries by the colony hybridization possessed a length of about 1800 bp. Since the potential poly(A) addition signals can be found at nucleotide position 1831 to 1836 (base Nos. 1831 to 1836) in the 3' noncoding region of pF58 cDNA (Fig. 2), the two different mRNAs for human Fas antigen, found by the northern hybridization, are probably generated by an alternative use of two different poly(A) addition signals.

According to the present invention, the cDNA coding for the human Fas is cloned and the nucleotide sequence is clarified. For people skilled in the art, therefore, it pertains within the scope of the present invention to construct an expression vector capable of expressing a recombinant Fas antigen in a suitable host system. Then, by transforming the host cells with the thus constructed expression vector, the transformed cells are cultured under the conditions suitable for expressing the DNA encoding the Fas antigen in order to prepare a recombinant human Fas antigen. The thus obtained recombinant human Fas antigen is useful in clarifying the apoptosis mechanism of various cells such as immune system cells, and is further effective in preparing monoclonal antibodies that specifically react with tumor cells expressing Fas or of value for the study, research and clinical test of those related to cytolytic activity of TNF.

For instance, the analysis of the cDNA coding for the human Fas antigen as obtained in Example 1 and the analysis of the corresponding encoded amino acid sequences, indicate that the Fas antigen belongs to a group of cell surface receptor proteins.

Here, the proteins thus provided include ones that may be encoded by the DNA of the present invention and may be defined to be the human Fas antigen and the functional homologs thereof. They may be cell surface proteins that are recognized by a monoclonal antibody capable of specifically recognizing the human Fas antigen and that induce apoptosis in the cells with the antibody alone without the presence of any other cytotoxic factor such as complement and the like. Particularly, the present invention provides proteins having the amino acid sequence disclosed in Figures 1 and 2 or peptides which are a part of the amino acid sequences thereof.

With the current technical level in this field of science, it will be easily contrived to introduce mutation such as deletions, additions, insertions and/or substitutions to the amino acid sequence without changing fundamental properties (e.g. physical properties, physiological or biological activity, immunological activity, etc.) of the proteins. For instance, substitution of a hydrophobic amino acid residue with other hydrophobic amino acid residue, or of amino acid residue having positive electric charge with other amino acid residue having positive electric charge, mutual substitution among Glu and Asp or Lys, His and Arg, substitution among Ile, Val, Met and leu groups, substitution among Gly, Ala, Ser and Cys groups, and substitution among Trp, Tyr and Phe groups may be predicted. For easy purification of the proteins of the present invention, furthermore, other proteins such as β -galactosidase of *Escherichia coli* or mouse IgG Fc fragment may be added to the N-terminal side or/and the C-terminal side of the proteins by the genetic engineering method, or the amino acid sequence may be partly cleaved or substituted by the similar method in order to more deeply analyze the function of the proteins, as can be easily contrived by people skilled in the art. Therefore, such human Fas antigen amino acid mutants are also encompassed by the present invention. For instance, soluble Fas antigens indicated by amino acid Nos.1 to 157 are preferred examples of such mutants.

The nucleotide sequences of cDNAs coding for the human Fas antigen of the present invention are shown in Figs. 1 and 2. It would be understood that Fas derivatives having substantially the same functions as the natural Fas antigen determinant can be obtained from the above DNAs by inserting, deleting, substituting or cleaving the nucleotides. Therefore, the DNAs thus derived are also encompassed by the scope of the present invention.

The insertion, substitution or deletion of the nucleotides can be carried out by, for example, the site directed mutagenesis, homologous recombination, cleavage with restriction enzymes, or ligation with ligase. The above methods can further be suitably combined with the primer extension using synthetic DNA fragments as primers or the polymerase chain reaction. These methods can be carried out in compliance with the methods disclosed in, for example, Sambrook et al. "Molecular Cloning, A Laboratory Manual, 2nd edition, Cold Spring Harbor Laboratory, 1989, Muramatsu (Ed.) "Labomannual Genetic Engineering" Maruzen

Co., 1988, Erlich HE, (Ed.) [PCR Technology, Principle of DNA Amplification and Its Application] Stockton Press, 1989, or in compliance with the modified methods thereof.

In the technical field of genetic engineering, furthermore, it has been known to substitute the bases in the base sequence for other base sequence without changing the amino acid sequence that is encoded thereby. Most of the amino acids are encoded by a plurality of genetic codes. For instance, Val is encoded by any one of GTT, GTA, GTC or GTG and Ala is encoded by any one of GCA, GCT, GCC or GCG. Therefore, the genetic base sequences of the present invention include base sequence substituted mutants that accompany the degeneracy of genetic codes.

From the disclosure of the present invention, furthermore, it would be easy in the art to add a base sequence such as a promoter or an enhancer to the 5' end side in order to produce a large amount of protein encoded by the DNA base sequence, in a transformant, to add a poly A addition signal base sequence to the 3' end side in order to stabilize the mRNA after the transcription, and/or to remove bases from or insert bases in the base sequence of the present invention in order to obtain mutant proteins from which amino acids are partly removed or to which amino acids are partly added in an attempt to further extensively analyze the function of the proteins encoded by the base sequence of the present invention. Therefore, the present invention further encompasses the base sequences having one or more bases that are added, altered, removed or inserted on the 5' end side or on the 3' end side and/or between them in the base sequence of the present invention.

The DNAs of the present invention include DNAs complementary to the DNAs encoding Fas or their fragments, DNAs capable of hybridizing with DNAs which are complementary to the DNAs encoding Fas or their fragments, and DNAs capable of hybridizing with human Fas protein cDNA fragments.

The expression vectors containing DNA coding for the human Fas antigen of the present invention can be constructed by methods known in the art. The vector suitable for expressing human Fas antigen DNA may have a promoter for initiating transcription closely on the upstream side of the DNA inserted site. Suitable promoters have been known in the art and can be selected by depending upon the functional characteristics in the host cells. Examples include a promoter of SV40 virus early gene, promoter of peptide chain elongation factor EF-1 α , promoter of metallothioneine gene, promoter of β -actin, and promoter of CMV virus that can be used for the expression in the animal cell systems, as well as a promoter of T7 polymerase and promoter of β -galactosidase gene that can be used for the expression in bacteria, particularly *Escherichia coli*, and promoters of phosphoglyceraldehyde dehydrogenase and alcohol dehydrogenase that can be used for the expression in yeasts. It is desired that a termination signal exists at a position downstream of a human Fas DNA inserted site.

In the case of animal cells, such regulators may be those from the human Fas sequence or from other sources of genes. When *Escherichia coli* is used, however, such regulators should desirably be from the *Escherichia coli* gene.

It is desired that the vector comprises a marker for selection such as a drug-resistant marker. A particularly desired example of the marker may include a neomycin-resistance gene, etc. an expression vector containing Fas DNA and a plasmid coding for drug resistance such as an antibiotic may be subjected to the transformation simultaneously.

In order to construct the expression vector, the DNA coding for the human Fas of the present invention is inserted in a suitable vector which can be selected from those already known in the art by taking into consideration of the promoters, termination signal, selection marker and other conditions. Examples of the DNA vector in which the cDNA of the invention is inserted and which is introduced into the host culture cells for expression the cDNA include pKCR, pEF-BOS, CDM8, pCEV4, bovine papilloma virus DNA for expression in the animal cells, pGEMEX, pUC, etc. for expression in *Escherichia coli*, as well as pY100 YCpAD1, etc. for expression in the yeasts.

Any culture cells may be used for the expression of human Fas antigen of the present invention as long as they are self-replicable and are capable of expressing the DNAs of the present invention. Examples include procaryotic microorganisms such as *Escherichia coli* and eucaryotic microorganisms such as yeasts (*Saccharomyces*, such as *S.cerevisiae*), as well as tissue culture cell lines derived from eucaryotic living things. Examples of *Escherichia coli* strains suitable for hosts include HB101, DH1, x1776, JM101, and JM109 of which the transformants can be easily sorted depending upon their resistance against drugs and enzymatic activities. Tissue culture cell lines include culture cells derived from insects, birds, mouse, rat, hamster, ape and human. Preferred examples are L cells, 3T3 cells, FM3A cells, CHO cells, COS cells, Vero cells, Hela cells and primary-cultured fibroblasts. Suitable host-vector systems and their use have been known in the art. Among them, any systems can be arbitrarily selected as long as they are suitable for expressing the DNAs of the present invention.

The proteins of the present invention can be produced in such a system by cultivating a host

(transformant) under the conditions suitable for the growth and capable of functioning the promoter of vector possessed by the host. These conditions can also be suitably selected and put into practice by people skilled in the art.

The present invention will be described more concretely by the following examples, but they should not be interpreted as limiting the invention in any manner.

In the specification, the technical terms, abbreviations and symbols are those which are conventionally used in the art unless otherwise stated. Moreover, the processes were conducted by making reference to Sambrook et al. "Molecular Cloning, A Laboratory Manual, 2nd edition", Cold Spring Harbor Laboratory, 1989, Imai Fumio et al., "Introduction of Recombinant Gene into Cells and Expression", Proteins, Nucleic Acids, Enzymes, Special Edition 28 (14), 1983, Yoshio Okada, "Summary of Cellular Engineering Technology", Experimental Medicine, Special Edition 7 (13), 1989, etc.

Example 1 Cloning of cDNA encoding human Fas

(1) Cell and antibody

Human lymphoma cell lines KT-3 (8×10^4 , kindly provided by Dr. Shimizu, Kanazawa Medical University) were grown in PRMI 1640 medium supplemented with 10% fetal calf serum (FCS) and 5 ng/ml human recombinant IL-6 (kindly provided by Ajinomoto Co., Inc.). The cell culture (total volume: 2 l) was incubated at 37 °C for 2 days under 5% CO₂-95% air.

Mouse T cell lymphoma WR19L cells (ATCC TIB52) (kindly provided by Dr. T. Kinebuchi, Tokyo Institute for Immunopharmacology, Inc.) were grown in RPMI 1640 medium containing 10% FCS.

Monkey COS-7 cells (ATCC CRL1651) and mouse L929 cells were grown in Dulbecco's modified Eagle medium (DMEM) containing 10% FCS.

Mouse anti Fas monoclonal antibody (IgM) was prepared in the same manner as mentioned above [Yonehara et al. (1989) op. cit.] and purified by column chromatography on hydroxyapatite.

(2) Construction of cDNA Library

Total RNA (2.7 mg) was prepared from the KT-3 cells (1.2×10^9), by the guanidium isothiocyanate/acid phenol method [Chomczynski and Sacchi, Anal. Biochem., 162, 156-159 (1987)] and poly(A)RNA (137 µg) was selected by means of an oligo(dT)-cellulose column chromatography. The poly(A)RNA (5 µg) was employed in synthesis of cDNA. Double strand cDNA primed with random hexamer oligonucleotide (pdN₆) or oligo(dT) was synthesized in the same manner as described in the report [Fukunaga et al., Cell, 61: 341-350 (1990)] except that M-MLV RNaseH - reverse transcriptase was employed instead of the AMV reverse transcriptase.

After addition of BstXI non-palindromic adapter (2 µg), DNA ligase (350 units), and ATP (final concentration: 1.0 mM), the mixture was reacted at 4 °C for 18 hours to ligate the adaptors to both ends of the synthesized double stranded DNA. The cDNA larger than 2 kb was recovered from the agarose gel and 0.25 µg of the recovered cDNA was ligated to BstXI-digested mammalian expression vector pCEV4 (0.2 rig) [Itoh et al., Science, 247, 324-327 (1990)] to construct the cDNA library. E. coli VM1100 cells were transformed with the cDNA by the electroporation method [Dower et al., Nucleic Acids Res., 16, 6127-6145 (1988)]. The individual clones of about 4.3×10^5 from the oligo(dT)-primed cDNA library were mixed with the clones of about 4.0×10^5 from the random hexamer-primed cDNA library and transfection with COS-7 cells was carried out as described below to recover the cDNA clones.

(3) Recovery of cDNA by Panning

The panning plates (panning dishes) were prepared as described below.

The bacterial 6 cm dishes (plates) (Falcon 1007) were incubated at room temperature for 90 minutes with 3 ml of 50 mM Tris-HCl (pH 9.5) containing µg/ml goat anti-mouse IgM (Cappel). The plates were washed three times with 0.15M NaCl and then incubated at room temperature overnight with 3 ml of phosphate-buffered saline (PBS).

One hundred and eight 6 cm dishes each containing 50% confluent monkey COS-7 cells (ATCC CRL1651), which were incubated in Dulbecco's modified Eagle medium containing 10% FCS, were transfected by the spheroplast fusion method [Sandri-Goldrin et al., Mol. Cell. Biol., 1, 743-752 (1981)] using the KT3 cDNA library comprising about 8×10^5 individual clones as described above.

After 72 hours from the transfection, the cells were detached from the dishes by incubation in PBS

containing 0.5 mM EDTA and 0.02% NaN_3 (PBS/EDTA/ NaN_3) at 37 °C for 30 minutes. The detached cells were pooled, collected by centrifugation and then suspended in 9 ml of cold PBS/EDTA/ NaN_3 containing 10 $\mu\text{g}/\text{ml}$ anti-Fas antibody. After incubation on ice for 60 minutes, the cells were diluted with an equal amount of PBS/EDTA/ NaN_3 and centrifuged at 1000 rpm for 5 minutes through PBS/EDTA/ NaN_3 containing 2% FicolI 400. The pelleted cells were resuspended in 27 ml of PBS/EDTA/ NaN_3 supplemented with 5% FCS and filtrated through Nylon meshes (pore size of 100 μm) to remove the aggregates. Then, the cells were distributed into 54 panning plates, each containing 5 ml of PBS/EDTA/ NaN_3 and 5% FCS. After incubation at room temperature for 2 to 3 hours, the Fas-expressing cells were adhered onto the plates and then nonadhering cells were removed by gently washing three times with 2 ml of PBS/EDTA/ NaN_3 containing 5% FCS. Then, the extrachromosomal DNA was prepared from the adhered COS cells according to the Hirt method [(1967), op. cit.]. More specifically, into each plate was placed 0.4 ml of 0.6% SDS solution containing 10 mM EDTA and each plate was incubated at room temperature for 20 minutes. The lysates were collected into microfuge tubes, NaCl was added up to 1M and the tubes were placed on ice for at least 5 hours. After centrifuged at 13,000 rpm, for 5 minutes, the supernatants were extracted with phenol/chloroform and the DNA was recovered by ethanol precipitation. With the DNA recovered from the first round of panning was transformed *Escherichia coli* VM1100 to give about 3.2×10^5 colonies. They were subjected to spheroplast fusion with COS cells in 48 plates, each being of 6 cm. Panning was performed with 24 plates in the same manner as described above and the DNA was prepared from the adhered cells. The so recovered DNA was transformed to give about 10,000 colonies, which were used for the third cycle of the spheroplast fusion with COS cells (24 plates, each being of 6 cm) and panning was performed in 12 plates, each being of 6 cm, to prepare the DNA from the adhered cells.

Transformation of *E. coli* VM100 was performed with the DNA finally obtained by the said three procedures and, among 2.8×10^5 clones, 14 of the resultant clones (pF1-pF14) were analyzed.

By digestion of the 14 plasmid DNA's with restriction enzyme, it has been elucidated that one group has the same insert of 3.0 kb (pF1, 2, 5, 11), while another group has the same insert of 1.5 kb (pF3, 4, 6, 7, 9).

By using the pF1 and pF3 among them, COS cells were subjected to transfection and the cells were analyzed by a flow cytometry using anti-Fas antigen to confirm the two cDNAs code for Fas antigen determinant.

The restriction mapping and DNA sequence analysis of pF1 and pF3 showed that they share identical sequences at the 5' end up to 0.57 kb, but their sequences at the 3' end diverge completely.

Then, the cDNA libraries of the above-mentioned KT-3 cells were screened by colony hybridization using the XhoI-BamHI DNA fragment (about 520 bp) as the 5' end of pF3. Ten colonies were obtained from 2×10^5 clones, said 10 clones showing identical restriction maps and overlapped each other. The longest cDNA clone was selected and designated pF58. Schematic representations and restriction maps of the pF58 and the said pF1 and pF3 are shown in Fig. 3A. In the Fig. 3A, the open box represents the open reading frame, the hatched box represents the signal sequence, and the black box represents the transmembrane region, respectively. In the representations for pF1 and pF3, the solid lines show identical sequence to that of pF58, while the dotted lines show difference sequence from that of pF58. However, the pF3 cDNA contains a single base (T) deletion at the position indicated with an arrowhead, the point of which is different from the pF58 cDNA.

Fig. 3B shows a hydropathy plot of human Fas antigen, which was obtained by the method of Kite and Doolittle [J. Mol. Biol., 157, 105-132 (1982)]. The numbers under the plot show positions of the amino acid residues of the precursor protein.

Then, the nucleotide sequence of the clone pF58 and its predicted amino acid sequence were determined. The results are shown in Fig. 1 and Fig. 2.

The cDNA analysis has elucidated the following points:

(1) The cDNA consists of 2534 bp and has a poly(A) addition signal (ATTAAA) at the 3'-end.

(2) There is a long open reading frame (1,008 nucleotides). The open reading frame can code for a protein consisting of 335 amino acids, starting from the initiation codon at the nucleotide positions 195 to 197 and ending at the termination codon TAG at the positions 1200 to 1202.

The results of the hydropathy analysis of the amino acid sequence suggested the presence of a signal sequence at the N-terminal end (See, Fig. 3B). Comparison with typical signal peptide cleavage sites suggested that the mature protein start at the 17th amino acid (Arg).

Therefore, the mature Fas antigen is a protein consisting of 319 amino acids with a calculated molecular weight of 36,000 and has the transmembrane segment consisting of 17 uncharged amino acids from Leu-154 to Val-170. And, it is followed by 3 basic amino acids at the cytoplasmic domain, as observed in other membrane-spanning proteins.

It has been indicated from the above results that this protein consists of an extracellular domain of 157 amino acids, a membrane-spanning domain of 17 amino acids and a cytoplasmic domain of 145 amino acids and that the extracellular domain is rich in cysteine residue (18 residues in 153 amino acids) and the cytoplasmic domain is relatively abundant in charged amino acids (24 basic amino acids and 19 acidic amino acids in 143 amino acids).

In Fig. 1 and Fig. 2 showing the nucleotide sequence and amino acid sequence of the Fas protein, the numbers above and below each line refer to the nucleotide position and the amino acid position, respectively. Amino acid numbers start at Arg-1 of the mature Fas protein. The transmembrane domain is underlined and two potential N-linked glycosylation sites (Asn-X-Ser/Thr) are indicated by asterisks. Three poly(A) addition signals (ATTAAA) are indicated as overlined. The nucleotide deleted in the pF3 is indicated with an arrowhead.

(3) Comparison in sequences of the Fas antigen with other members of the NGFR/TNFR family.

Comparison of the amino acid sequence of the Fas antigen with the sequences of other members of the NGFR/TNFR family was performed. The results are shown in Figs. 7 ~9.

Fig. 7 is a schematic representation of the cysteine-rich repeats of the extracellular domain. In open boxes, the cysteines are represented with bars, and the stripped boxes in the cytoplasmic domain represent the conserved region among the Fas antigen, the TNF receptor type I and the CD40 antigen. It has been indicated from this Fig. that the extracellular domains of the TNF receptor, the NGF receptor and the CD40 antigen can be divided into 4 cysteine-rich subdomains, while the Fas antigen and the CD40 antigen contain 3 subdomains.

Fig. 8 shows the amino acid sequences of the extracellular domains of human Fas (hFas), human TNF receptor type I (hTNFR1) [Schall et al., 1990], human TNF receptor type II (hTNFR2) [Smith et al., Cell, 61, 361-370 (1990)], human NGF receptor (hNGFR) [Johnson et al., Science, 248, 1019-1023 (1986)], human CD40 (hCD40) [Stamenkovic et al., EMBOJ., 8, 1403-1410 (1989)] and rat OX40 (rOX40) [Mallett et al., EMBO J., 9, 1063-1068 (1990)]. Gaps(-) are introduced to optimize matches. Identical amino acids are boxed.

It has been indicated from this Fig. that the positions of the cysteine residues are well conserved. The numbers referring to residues are followed as in references. The amino acid residues conserved among the cysteine-rich repeating units are indicated at the bottom of the sequence. Fig. 9 is a comparison representation of the cytoplasmic domains of the Fas, the TNF receptor I and the CD40. The amino acid sequences of the corresponding regions of the hCD40, hFas and hTNFR1 are aligned. Identical and conserved amino acids are boxed in solid and dotted lines, respectively.

It has been established that the Fas of this invention belong to the group of such cell surface proteins.

Example 2 Preparation of transformants expressing Fas antigen

The 2.6 kb XhoI fragment containing the Fas cDNA was prepared from the plasmid pF58 (2 µg) and transfected into the BstXI site of a mammalian expression plasmid pEF-BOS [Mizushima and Nagata, Nucleic Acids Res., 18, 5322 (1990)] using a BstXI adapter to construct the expression vector pEFF58 containing the Fas-coding cDNA under the control of human peptide chain-elongation factor 1α gene.

(1) Transformation of mouse fibroblastoma L929 cells was performed according to the following method:

L929 cells 1×10^6 , which were grown in DMEM containing 10% FCS, were cotransfected with 0.2 µg of pStneoB containing neomycin-resistant genes and 20 µg of ApaI-digested pEFF58 in a 10 cm plate by the calcium phosphate coprecipitation method [Sambrook et al. "Molecular Cloning, A Laboratory Manual, 2nd edition", Cold Spring Harbor Laboratory, 1989], followed by treatment with glycerol. After 12 hours from the transfection, the cells were treated with trypsin, diluted ten times and neomycin-resistant cells were selected in a medium containing 0.4mg/ml G-418.

After sufficient growth, the cells were washed with PBS/EDTA/Na₃ containing 5% FCS and incubated for 60 minutes on ice in the same buffer containing 10 µg/ml mouse anti-Fas antigen. The expression of the Fas antigen in the transformants was examined by the following processes:

The cells were washed to remove the unbound anti-Fas antibody and then stained for 30 minutes on ice with 10 µg/ml FITC-conjugated goat anti-mouse IgM (Cappel). The cells were centrifuged at 1,000 rpm for 5 minutes through a cushion of PBS/EDTA/Na₃ containing 2% Ficoll, and analyzed on a FACScan (Becton Dickson Instruments, USA).

(2) Transformation of mouse T-cell lymphoma WR19L cells was performed by the following method:

WR19L cells (1×10^7 in 0.8 ml, ATCC TIB52, kindly provided by Dr. T. Kinebuchi, Tokyo Institute for Immunopharmacology, Inc.), which were grown in RPMI1640 containing 10% FCS, were cotransfected with 2.5 μ g/ml EcoRI-digested pMAMneo (Clontech) and 25 μ g/ml VspI-digested pEFF58 by electroporation [Potter et al., Proc. Natl. Acad. Sci. USA, 81, 7161-7165 (1984)] [at 290V, with a capacitance of 950 μ F; Gene Pulser (Bio-Rad)]. The cells were cultured in a growth medium in 96-well microtiter plates (0.1ml/well) for 2 days and neomycin-resistant clones were selected in a medium containing G-418 at a final concentration of 900 μ g/ml. After 9 days, the expression of the Fas antigen in individual G-418-resistant transformants was analyzed on a flow cytofluorometer by mouse anti-Fas antibody and the Fas-positive cells were cloned by a limiting dilution method. Then, the WR19L transformant clone, F58-12a, expressing the Fas antigen was analyzed by a Western Blotting method.

(3) Western Blotting of F58-12a

Membrane fractions from the mouse WR12L cell line, its transformant clone expressing the Fas antigen (58-12a) and human KT-3 were analyzed by Western Blotting with anti-Fas antibody or control IgM. The results showed a specific band with an apparent molecular weight of 43,000. This value is in good agreement with that calculated from the Fas antigen amino acid sequence, in considering the difference wherein the sugar moieties may be attached to the two potential N-glycosylation sites on the extracellular domain of the Fas antigen as shown in Fig. 2.

Experimental Example 1 Cytolytic activity of anti-Fas antibody on Fas-expressing cells

As described hereinabove, mouse anti-Fas monoclonal antibody showed a cytolytic effect on human cells (U-937, HL-60, A637 or FL cells), but the antibody does not react with mouse cells [Yonehara et al., op. cit.].

In this Example, it was examined whether the polypeptide coded by the present pF58 cDNA may mediate the cytolytic activity of anti-Fas antibody. Mouse WR19L and mouse L929 were transformed as described in Example 2 to prepare transformant cells expressing Fas antigen. These cells are different in the point wherein L929 cells can be killed by TNF in the presence of actinomycin D, while WR19L cells are susceptible to the cytolytic activity of TNF in the presence or absence of any metabolic inhibitors.

As described hereinabove, the expression plasmid pEFF-58 and a plasmid carrying the neo-resistance gene were cotransfected into WR19L cells or L929 cells and selection in the presence of G-418 afforded several G-418-resistant clones.

Then, parental WR19L and L929 cells, 2 transformants derived from WR19L (58-12a and 58-80d) and 2 clones derived from L929 (LB1 and LB11) were stained with anti-Fas antibody (IgM) and anti-mouse IgM antibody bound with FITC, followed by subjecting to flow cytofluorometry.

The results are shown in Fig. 4, wherein A: WR19L; B: 58-12a; C: 58-80d; D: L929; E: LB1; F: LB11.

As apparent from the Fig. 4, the parental cells, mouse WR19L and L929 cells, did not express the Fas antigen, while the WR19L transformant cells (58-12a, F58-80d) and L929 cells (LB1 and LB11) extremely abundantly expressed the Fas antigen on their surfaces.

Then, the cytolytic effect of the Fas antibody was examined using the Fas antigen-expressing cells.

The mouse WR19L cell and its transformant clones (58-12a and 58-80d) were incubated with various concentrations of anti-Fas antibody (0~1 μ g/ml) at 37°C for 24 hours. Viable and dead cell counts were determined by the trypan blue exclusion method. The results are shown in Fig. 5, wherein open squares represent WR19L, closed circles represent 58-12a and closed squares represent 58-80d. As apparent from the Fig. 5, the F58-12a and F58-80d cell lines responded to the anti-Fas antibody in a concentration-dependent manner. The half-maximal response was obtained at 0.1 μ g/ml concentration of the anti-Fas antibody and the cells were completely killed by incubation for 24 hours in the presence of 1 μ g/ml said antibody.

The cytolytic effect of the anti-Fas antibody on the L929 transformant clones was examined according to the following method.

The L929 cells and the transformant clones expressing recombinant human Fas antigen (LB1 and LB11) were dispersed onto 96-well microtiter plates (25,000 cells/well) and incubated for 24 hours. Actinomycin D was added at a final concentration of 0.5 μ g/ml and the cells were incubated with various concentrations of anti-Fas

antibody (30ng~2 μ g/ml) at 37°C for 17 hours. Then, the cells were stained with a solution of 0.75% crystal violet in 50% ethanol, 0.25% NaCl and 1.75% formaldehyde at room temperature for 20 minutes. Dye uptake was assessed by the OD value measured at 540 nm using Micro-ELISA autoreader, as expressed as

a percentage of the OD measured value without anti-Fas antibody. The results are shown in Fig. 6, wherein open squares represent L929, closed circles represent LB1 and closed squares represent LB11.

As apparent from the Fig. 6, the LB11 and LB1 cell lines responded to the anti-Fas antibody in the presence of actinomycin D in a similar concentration-dependent manner to that of the WR19L cells expressing Fas.

In any cases, the parental mouse WR19L and L929 were not affected by the anti-Fas antibody at a concentration of 1 μ g/ml under the same conditions.

Experimental Example 2 Apoptosis induced by anti-Fas antibody

Apoptosis of cells induced by Fas was proved according to the following method:

(1) Fragmentation of Chromosomal DNA

The WR19L cell and its transformant clones, 58-12a and 58-80d cells, were incubated in the presence of 300ng/ml anti-Fas antibody or 60ng/ml mouse TNF- α . Before incubation and after 1 hour, 2 hours and 3 hours incubation, total DNA was prepared from cells and analyzed by 2% agarose gel electrophoresis in the presence of 0.5 μ g/ml ethidium bromide. The fragmentation of chromosomal DNA was observed. The fragmented DNA was separated in a ladder pattern and its minimum size was approximately 180 bp. This ladder DNA fragments were observed within 1 hour of incubation and more than 60% of chromosomal DNA was fragmented after 3 hours of incubation. On the other hand, the chromosomal DNA from the parental WR19L cells remained as a high molecular weight form even after incubation with the anti-Fas antibody.

A similar DNA fragmentation was observed in the parental WR19L cells and their transformant cells treated with 60 ng/ml TNF. This was similarly observed in L929 cells.

These results suggest that the specific binding of the Fas antibody to the Fas antigen on the cell surface induces an endonuclease which digests the chromosomal DNA. They are consistent with those properties of apoptosis observed in various systems [Schmid et al., (1987); Ucker, (1987); Smith et al., (1989); Williams et al, (1990), op. cit.]. And, the expression of the Fas antigen in mouse WR19L and L929 cells does not affect a cell-killing effect of TNF and the transformant cells were also killed with mouse TNF- α at the same concentration as in parental cells.

(2) Morphological changes

Morphological changes in the L929 transformant expressing the Fas antigen were examined.

Morphological changes of the LB1 cells were initiated after incubation in the presence of 0.5 μ g/ml actinomycin D and in the presence of 1 μ g/ml anti-Fas antibody for 3 hours and, after 5 hours, many typical apoptotic blebs were seen on cell surface. Then, almost all cells were detached from plates within 24 hours. Such morphological changes of the LB1 cells were not observed even in the presence of actinomycin D unless the Fas antibody was present. And, the anti-Fas antibody did not give any morphological changes to parental L929 cells.

It becomes apparent, as described in the above Experimental Example, that the human Fas antigen obtained in this invention can mediate apoptosis of cells. Recombinant human Fas can be prepared using the present cDNA by a recombinant DNA technology. Further, the monoclonal antibody to specifically act the human Fas can be also prepared readily in a well-known manner. Thus, these are provided diagnostic and therapeutic means for diseases and disorders in which the cells expressing the Fas antigen would participate.

SEQUENCE LISTING

5 SEQ ID NO:1
 SEQUENCE TYPE: nucleic acid
 SEQUENCE LENGTH: 2534 base pairs

 STRANDEDNESS: double
 TOPOLOGY: linear
 MOLECULE TYPE: cDNA to mRNA
 10 ORIGINAL SOURCE
 ORGANISM: Homo sapiens
 IMMEDIATE SOURCE
 LIBRARY: pCEV4
 CLONE: clone pF58
 15 FEATURES:
 from 195 to 1202 bp CDS
 IDENTIFICATION METHOD: S

 from 195 to 242 bp signal peptide
 IDENTIFICATION METHOD: S
 20 from 243 to 1199 bp mature peptide
 IDENTIFICATION METHOD: S

 from 1831 to 1836 bp polyA site
 IDENTIFICATION METHOD: S
 25 from 2352 to 2357 bp polyA site
 IDENTIFICATION METHOD: S

 from 2518 to 2523 bp polyA site
 IDENTIFICATION METHOD: S
 30 GACGCTTCTG GGGAGTGAGG GAAGCGGTTT ACGAGTGACT TGGCTGGAGC CTCAGGGGCG 60
 CGCACTGGCA CGGAACACAC CCTGAGGCCA GCCCTGGCTG CCCAGGCGGA GCTGCCTCTT 120
 CTCCCGCGGG TTGGTGGACC CGCTCAGTAC GGAGTTGGGG AAGCTCTTTC ACTTCGGAGG 180
 35 ATTGCTCAAC AACC ATG CTG GGC ATC TGG ACC CTC CTA CCT CTG GTT CTT 230
 Met Leu Gly Ile Trp Thr Leu Leu Pro Leu Val Leu
 -16 -15 -10 -5
 ACG TCT GTT GCT AGA TTA TCG TCC AAA AGT GTT AAT GCC CAA GTG ACT 278
 Thr Ser Val Ala Arg Leu Ser Ser Lys Ser Val Asn Ala Gln Val Thr
 40 1 5 10
 GAC ATC AAC TCC AAG GGA TTG GAA TTG AGG AAG ACT GTT ACT ACA GTT 326
 Asp Ile Asn Ser Lys Gly Leu Glu Leu Arg Lys Thr Val Thr Thr Val
 15 20 25
 45 GAG ACT CAG AAC TTG GAA GGC CTG CAT CAT GAT GGC CAA TTC TGC CAT 374
 Glu Thr Gln Asn Leu Glu Gly Leu His His Asp Gly Gln Phe Cys His
 30 35 40

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| | | |
|----|---|------|
| | AAG CCC TGT CCT CCA GGT GAA AGG AAA GCT AGG GAC TGC ACA GTC AAT | 422 |
| | Lys Pro Cys Pro Pro Gly Glu Arg Lys Ala Arg Asp Cys Thr Val Asn | |
| | 45 50 55 60 | |
| 5 | GGG GAT GAA CCA GAC TGC GTG CCC TGC CAA GAA GGG AAG GAG TAC ACA | 470 |
| | Gly Asp Glu Pro Asp Cys Val Pro Cys Gln Glu Gly Lys Glu Tyr Thr | |
| | 65 70 75 | |
| | GAC AAA GCC CAT TTT TCT TCC AAA TGC AGA AGA TGT AGA TTG TGT GAT | 518 |
| | Asp Lys Ala His Phe Ser Ser Lys Cys Arg Arg Cys Arg Leu Cys Asp | |
| 10 | 80 85 90 | |
| | GAA GGA CAT GGC TTA GAA GTG GAA ATA AAC TGC ACC CGG ACC CAG AAT | 566 |
| | Glu Gly His Gly Leu Glu Val Ile Asn Cys Thr Arg Thr Gln Asn | |
| | 95 100 105 | |
| 15 | ACC AAG TGC AGA TGT AAA CCA AAC TTT TTT TGT AAC TCT ACT GTA TGT | 614 |
| | Thr Lys Cys Arg Cys Lys Pro Asn Phe Phe Cys Asn Ser Thr Val Cys | |
| | 110 115 120 | |
| | GAA CAC TGT GAC CCT TGC ACC AAA TGT GAA CAT GGA ATC ATC AAG GAA | 662 |
| | Glu His Cys Asp Pro Cys Thr Lys Cys Glu His Gly Ile Ile Lys Glu | |
| | 125 130 135 140 | |
| 20 | TGC ACA CTC ACC AGC AAC ACC AAG TGC AAA GAG GAA GGA TCC AGA TCT | 710 |
| | Cys Thr Leu Thr Ser Asn Thr Lys Cys Glu Glu Gly Ser Arg Ser | |
| | 145 150 155 | |
| 25 | AAC TTG GGG TGG CTT TGT CTT CTT CTT TTG CCA ATT CCA CTA ATT GTT | 758 |
| | Asn Leu Gly Trp Leu Cys Leu Leu Leu Pro Ile Pro Leu Ile Val | |
| | 160 165 170 | |
| | TGG GTG AAG AGA AAG GAA GTA CAG AAA ACA TGC AGA AAG CAC AGA AAG | 806 |
| | Trp Val Lys Arg Lys Glu Val Gln Lys Thr Cys Arg Lys His Arg Lys | |
| | 175 180 185 | |
| 30 | GAA AAC CAA GGT TCT CAT GAA TCT CCA ACC TTA AAT CCT GAA ACA GTG | 854 |
| | Glu Asn Gln Gly Ser His Glu Ser Pro Thr Leu Asn Pro Glu Thr Val | |
| | 190 195 200 | |
| 35 | GCA ATA AAT TTA TCT GAT GTT GAC TTG AGT AAA TAT ATC ACC ACT ATT | 902 |
| | Ala Ile Asn Leu Ser Asp Val Asp Leu Ser Lys Tyr Ile Thr Thr Ile | |
| | 205 210 215 220 | |
| | GCT GGA GTC ATG ACA CTA AGT CAA GTT AAA GGC TTT GTT CGA AAG AAT | 950 |
| | Ala Gly Val Met Thr Leu Ser Gln Val Lys Gly Phe Val Arg Lys Asn | |
| | 225 230 235 | |
| 40 | GGT GTC AAT GAA GCC AAA ATA GAT GAG ATC AAG AAT GAC AAT GTC CAA | 998 |
| | Gly Val Asn Glu Ala Lys Ile Asp Glu Ile Lys Asn Asp Asn Val Gln | |
| | 240 245 250 | |
| 45 | GAC ACA GCA GAA CAG AAA GTT CAA CTG CTT CGT AAT TGG CAT CAA CTT | 1046 |
| | Asp Thr Ala Glu Gln Lys Val Gln Leu Leu Arg Asn Trp His Gln Leu | |
| | 255 260 265 | |
| 50 | CAT GGA AAG AAA GAA GCG TAT GAC ACA TTG ATT AAA GAT CTC AAA AAA | 1094 |
| | His Gly Lys Lys Glu Ala Tyr Asp Thr Leu Ile Lys Asp Leu Lys Lys | |

| | 270 | 275 | 280 | |
|----|---|-----|-----|------|
| 5 | GCC AAT CTT TGT ACT CTT GCA GAG AAA ATT CAG ACT ATC ATC CTC AAG Ala Asn Leu Cys Thr Leu Ala Glu Lys Ile Gln Thr Ile Ile Leu Lys 285 290 295 300 | | | 1142 |
| | GAC ATT ACT AGT GAC TCA GAA AAT TCA AAC TTC AGA AAT GAA ATC CAA Asp Ile Thr Ser Asp Ser Glu Asn Ser Asn Phe Arg Asn Glu Ile Gln 305 310 315 | | | 1190 |
| 10 | AGC TTG GTC TAGAGTGAAA AACAACAAAT TCAGTTCTGA GTATATGCAA Ser Leu Val 320 | | | 1239 |
| | TTAGTGTGTTG AAAAGATTCT TAATAGCTGG CTGTAAATAC TGCTTGGTTT TTTACTGGGT | | | 1299 |
| 15 | ACATTTTATC ATTTATTAGC GCTGAAGAGC CAACATATTT GTAGATTTTT AATATCTCAT | | | 1359 |
| | CATTCTGCCT CCAAGGATGT TTAATCTA GTTGGGAAAA CAACTTCAT CAAGAGTAAA | | | 1419 |
| | TGCAGTGGCA TGCTAAGTAC CCAATAGGA GTGTATGCAG AGGATGAAAG ATTAAGATTA | | | 1479 |
| 20 | TGCTCTGGCA TCTAACATAT GATTCTGTAG TATGAATGTA ATCAGTGTAT GTTAGTACAA | | | 1539 |
| | ATGTCTATCC ACAGGCTAAC CCCACTCTAT GAATCAATAG AAGAAGCTAT GACCTTTTGC | | | 1599 |
| | TGAAATATCA GTTACTGAAC AGGCAGGCCA CTTTGCCCTCT AAATTACCTC TGATAATTCT | | | 1659 |
| 25 | AGAGATTTTA CCATATTTCT AAACTTTGTT TATAACTCTG AGAAGATCAT ATTTATGTAA | | | 1719 |
| | AGTATATGTA TTTGAGTGCA GAATTTAAAT AAGGCTCTAC CTCAAAGACC TTTGCACAGT | | | 1779 |
| | TTATTGGTGT CATATTATAC AATATTTCAA TTGTGAATTC ACATAGAAAA CATTAAATTA | | | 1839 |
| | TAATGTTTGA CTATTATATA TGTGTATGCA TTTTACTGGC TCAAACTAC CTACTTCTTT | | | 1899 |
| 30 | CTCAGGCATC AAAAGCATTT TGAGCAGGAG AGTATTACTA GAGCTTTGCC ACCTCTCCAT | | | 1959 |
| | TTTTGCCCTG GTGCTCATCT TAATGGCCTA ATGCACCCCC AATCATGGAA ATATCACCAA | | | 2019 |
| | AAAATACTTA ATAGTCCACC AAAAGGCAAG ACTGCCCTTA GAAATCTAG CCTGGTTTGG | | | 2079 |
| 35 | AGATACTAAC TGCTCTCAGA GAAAGTAGCT TTGTGACATG TCATGAACCC ATGTTTGCAA | | | 2139 |
| | TCAAAGATGA TAAATAGAT TCTTATTTTT CCCCCACCCC CGAAATGTT CAATAATGTC | | | 2199 |
| | CCATGTAAAA CCTGCTACAA ATGGCAGCTT ATACATAGCA ATGGTAAAAT CATCATCTGG | | | 2259 |
| 40 | ATTTAGGAAT TGCTCTTGTC ATACCCTCAA GTTTCTAAGA TTTAAGATTC TCCTTACTAC | | | 2319 |
| | TATCCTACGT TTAAATATCT TTGAAAGTTT GTATTAAATG TGAATTTTAA GAAATAATAT | | | 2379 |
| | TTATATTTCT GTAAATGTAA ACTGTGAAGA TAGTTATAAA CTGAAGCAGA TACCTGGAAC | | | 2439 |
| 45 | CACCTAAAGA ACTTCCATTT ATGGAGGATT TTTTGGCCCC TTGTGTTTGG AATTATAAAA | | | 2499 |
| | TATAGGTAAA AGTACGTAAT TAAATAATGT TTTTG | | | 2534 |

50 Claims

1. An isolated DNA coding for human cell surface antigen or analogue thereof having substantially the same function as said human cell surface antigen, a DNA analogue derived therefrom or a DNA fragmented therefrom.

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2. The DNA of claim 1, wherein the human cell surface antigen is a Fas antigen.

3. The DNA of claim 1 or 2, wherein the human cell surface antigen comprises at least a part of the amino

- acid sequences described in Figs. 1 and 2.
4. The DNA of claim 1 or 2, wherein the human cell surface antigen comprises the amino acid sequences described in Figs. 1 and 2.
 5. The DNA of any one of claims 1 to 4, wherein the DNA comprises at least a part of the base sequences described in Figs. 1 and 2.
 6. The DNA of any one of claims 1 to 5, wherein the DNA comprises the base sequences described in Figs. 1 and 2.
 7. The DNA of any one of claims 1 to 6, wherein the DNA comprises the base sequence from numbers 215 to 1199 of Figs. 1 and 2.
 8. The DNA of any one of claims 1 to 6, wherein the DNA is represented by the base sequence from numbers 243 to 1199 of Figs. 1 and 2.
 9. The DNA of any one of claims 1 to 6, wherein the DNA is represented by the base sequence from numbers 215 to 713 of Figs. 1 and 2.
 10. The DNA of any one of claims 1 to 6, wherein the DNA is represented by the base sequence from numbers 243 to 713 of Figs. 1 and 2.
 11. The DNA of any one of claims 1 to 6, wherein the DNA comprises at least a part of the base sequence from numbers 243 to 713 of Figs. 1 and 2.
 12. Human cell surface antigen, a protein which possesses substantially the same function as said antigen, or a peptide derived therefrom and having partial amino acid sequence thereof.
 13. The human cell surface antigen, protein or peptide of claim 12, wherein the human cell surface antigen is a Fas antigen.
 14. The human cell surface antigen, protein or peptide of claims 12 or 13, wherein said antigen, protein or peptide comprises at least a part of the amino acid sequences described in Figs. 1 and 2.
 15. The human cell surface antigen, protein or peptide of any one of claims 12 to 14, wherein said antigen, protein or peptide comprises the amino acid sequence from numbers -16 to 319 described in Figs. 1 and 2.
 16. The human cell surface antigen, protein or peptide of any one of claims 12 to 14, wherein said antigen, protein or peptide comprises the amino acid sequence from numbers 1 to 319 described in Figs. 1 and 2.
 17. The human cell surface antigen, protein or peptide of any one of claims 12 to 14, wherein said antigen, protein or peptide comprises the amino acid sequence from numbers -16 to 157 described in Figs. 1 and 2.
 18. The human cell surface antigen, protein or peptide of any one of claims 12 to 14, wherein said antigen, protein or peptide comprises the amino acid sequence from numbers 1 to 157 described in Figs. 1 and 2.
 19. The human cell surface antigen, protein or peptide of any one of claims 12 to 14, wherein said antigen, protein or peptide comprises at least a part of the amino acid sequence from numbers 1 to 157 described in Figs. 1 and 2.
 20. An expression vector comprising the DNA of any one of claims 1 to 11.
 21. A transformant transformed by the expression vector of claim 20.

22. The transformant of claim 21 wherein said transformant is derived from a procaryotic living cell.

23. The transformant of claim 21, wherein said transformant is Escherichia coli.

5 24. The transformant of claim 21, wherein said transformant is derived from a eucaryotic living cell.

25. The transformant of claim 21, wherein said transformant is an yeast.

26. The transformant of claim 21, wherein said transformant consists of mammalian culture cells.

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27. A method for producing the antigen, protein or peptide of any one of claims 12 to 19, which comprises culturing the transformant of any one of claims 21 to 26 under a suitable condition.

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28. A pharmaceutical composition containing the antigen, protein or peptide according to any one of claims 12 to 19, optionally in combination with a pharmaceutically acceptable carrier.

29. Use of the antigen, protein or peptide according to any one of claims 12 to 19 for the preparation of a pharmaceutical composition.

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30. Use of the antigen, protein or peptide according to any one of claims 12 to 19 as a reagent for analysis.

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Fig. 3

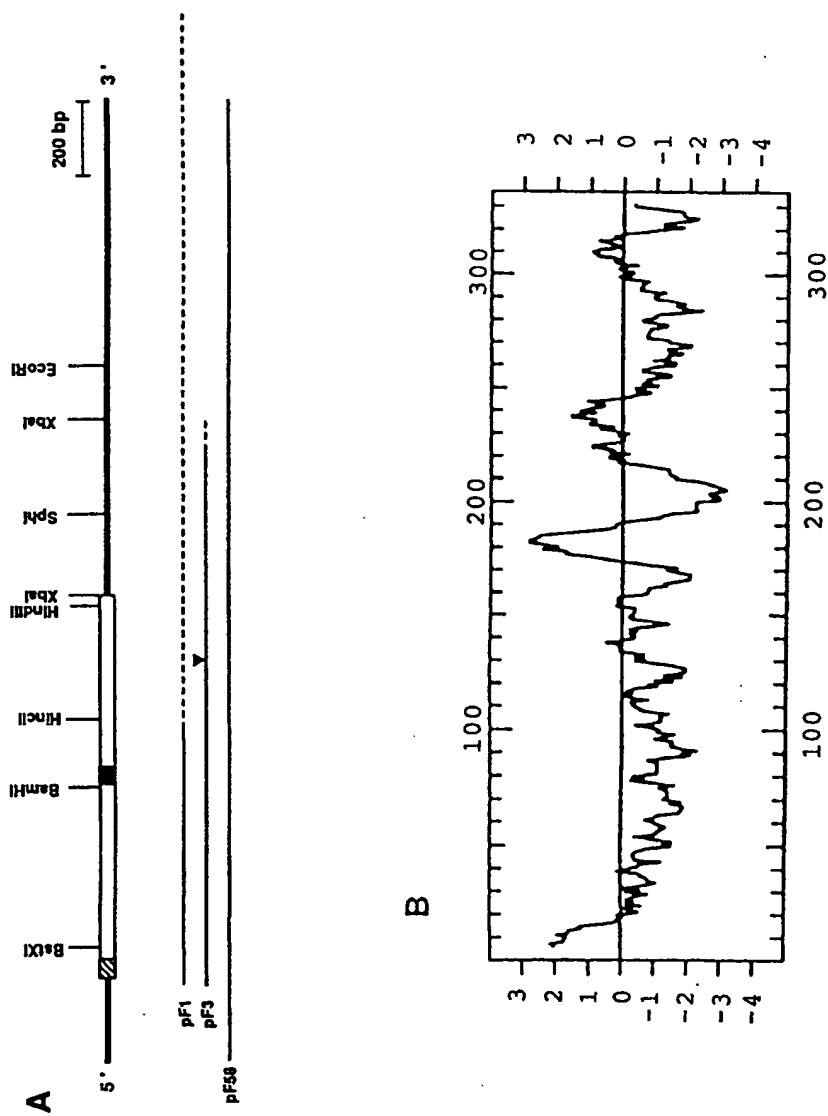


Fig. 4

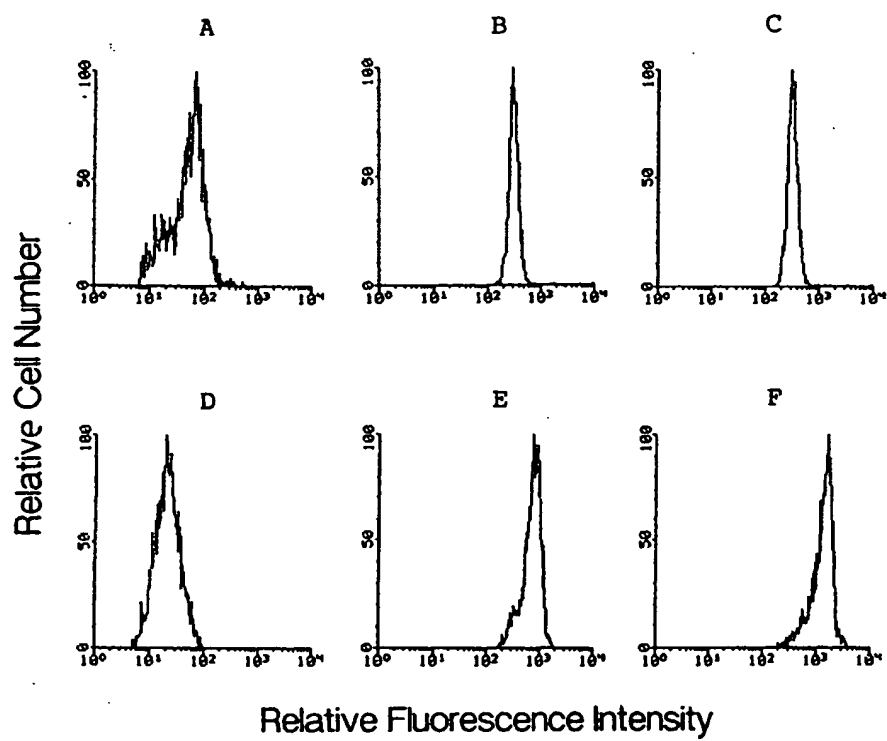


Fig. 5

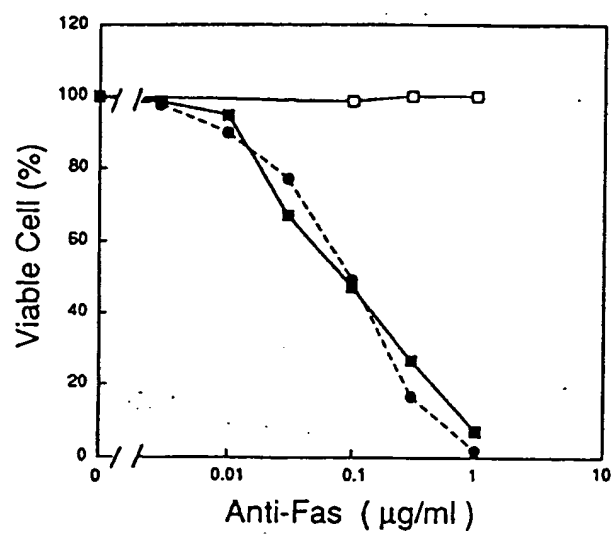


Fig. 6

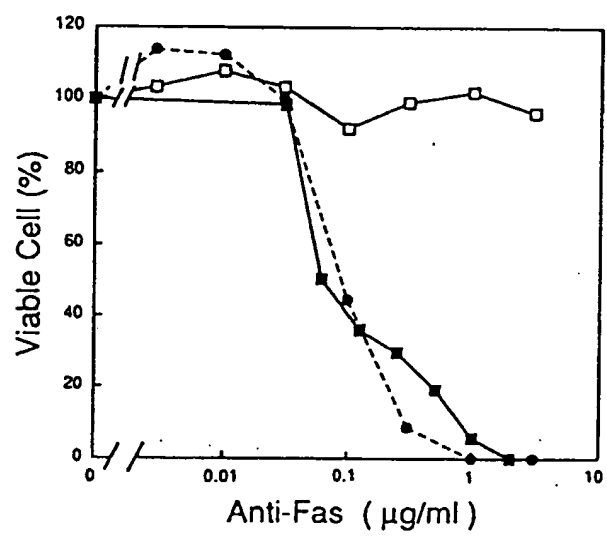


Fig. 7

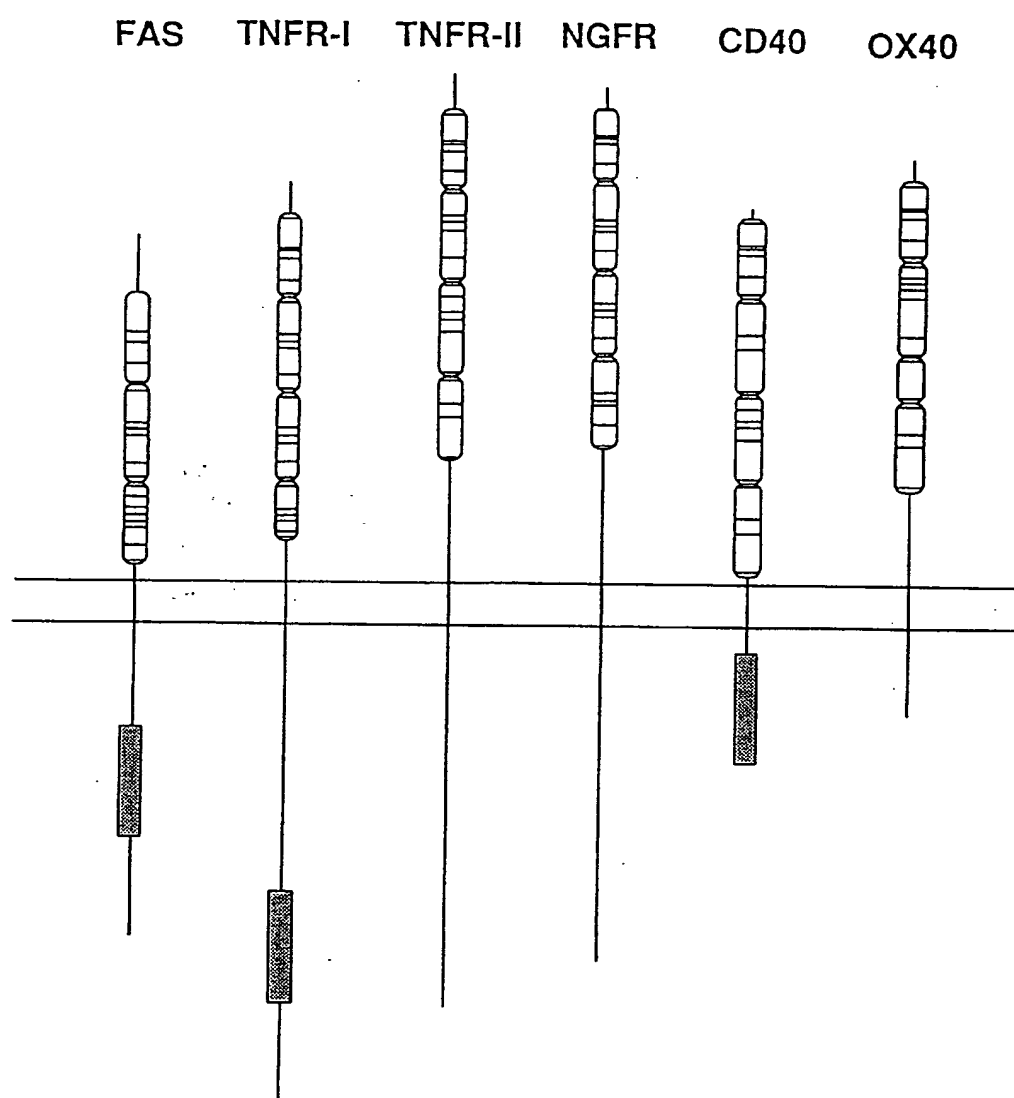


Fig. 9

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|------------------|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| hCD40 (225-269) | K | A | P | H | P | K | Q | E | P | Q | E | I | N | F | P | D | D | L | P | G | S | N | T | A | A | P | V | Q | E | T | L | H | G | C | Q | P | V | T | Q | E | D | G | - | K | E | S |
| hFAS (230-274) | K | G | F | V | R | K | N | G | V | N | E | A | K | I | - | D | E | I | K | N | D | N | V | Q | D | T | A | E | Q | K | V | Q | L | R | N | W | H | Q | L | H | G | K | K | E | A | |
| hTNFRI (332-376) | K | E | F | V | R | L | G | L | S | D | H | E | I | - | D | R | L | E | L | Q | N | G | R | C | L | R | E | A | Q | Y | S | M | L | A | T | W | R | R | T | P | R | R | E | A | | |



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Application Number

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| Y | | 2-11, 13-19 | |
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| Y | | 2-11, 13-19 | |
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| Place of search BERLIN | | Date of completion of the search 05 AUGUST 1992 | Examiner JULIA P. |
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| | | | TECHNICAL FIELDS SEARCHED (Int. Cl.5) |
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| The present search report has been drawn up for all claims | | | |
| Place of search BERLIN | | Date of completion of the search 05 AUGUST 1992 | Examiner JULIA P. |
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